

Central Australia Newsletter

April 2018

# From the Land for Wildlife Coordinator

The cover image is of the bark of a River Red Gum (Eucalyptus camaldulensis) beside the Todd River. This species has been voted the Eucalyptus of the year in 2018! In this issue of the newsletter we explore bark further—so read on!

"If we can get people excited about animals, then by crikey, it makes it a heck of a lot easier to save them."

— Steve Irwin

It seems to be butterfly season out there, with plenty of Caper White (Belenois java) emerging from their chrysalis on Wild Passionfruit (Capparis spinosa var nummularia). Have a wander to Araluen to check them out for yourself, or you may find them in your own backyard!

The hot weather finally seems to be easing off and so as it cools down you will see fewer reptiles so make the most of their presence and enjoy their company (thanks to all that have sent in images recently). Until next month...



A Caper White Butterfly (*Belenois java*) seen feeding on, pupating on, and breeding on a Wild Passionfruit (*Capparis spinosa var nummularia*) outside the Araluen Cultural Precinct in Alice Springs.

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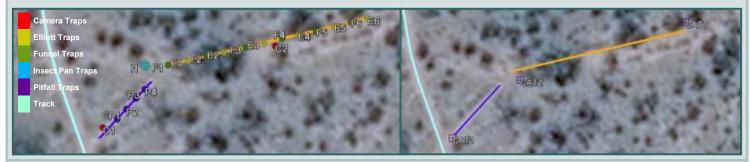
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# Biodiversity Survey with the Tjuwanpa Women Rangers

Land for Wildlife has been engaging with the Tjuwanpa Women Rangers for many years now, providing support for onground work and facilitating workshops with the Ntaria Junior Rangers. In early April, Land for Wildlife helped the Women Rangers to discover the wildlife in the region by conducting a short biodiversity survey, as well as assist with some feral Cat (*Felis catus*) trapping.

For the biodiversity survey, we used a range of techniques to assess the presence of wildlife (*see map to the left below*). This included the use of two camera traps, four pitfall traps, six Elliott traps, six funnel traps and an insect pan trap. We also smoothed out sand in the trap site to check for tracks each morning on arrival to the undisclosed site. The survey was conducted over two nights, though ideally biodiversity surveys would go for a longer period to ensure an adequate representation of the biodiversity of a region has been obtained. For the feral Cat trapping session, the Tjuwanpa Women Rangers set up three traps spaced along the ends of the biodiversity survey trap lines (*see map to the right below*) and trapped for two nights in line with the biodiversity survey.





Pitfall traps (top left) were established to capture small mammals, reptiles, invertebrates and other creatures. A netting fence is set up along a transect, with deep buckets set in the ground. The theory goes that if an animal crossing the sand bed comes across the net, it will follow it in an attempt to cross and eventually drop into a bucket. Funnel traps (top right) are designed with a large opening and a narrow end inside the mesh trap tube, with the ability to capture small reptiles, mammals and invertebrates. Elliott traps consist of an elongated box with a depression plate inside, designed to trap small mammals when baited with peanut butter and oats or curious lizards exploring the cave. An insect pan trap (bottom left) was set up in an open area and filled with water, salt and bio-friendly detergent. The camera traps (bottom right) are a method of recording biodiversity without actual capture of an individual, rather heat and movement sensors detect a passing animal and take several photographs as record of presence.

We were able to record the presence of three individual Desert Spadefoot Toads (*Notaden nichollsi*), one on the first morning and two on the second morning. These were quickly released from the pitfalls and upon release they were seen to burrow back into the sand for the day. One of the pitfalls was successful in trapping a Pie-dish Beetle (*Helea sp.*) and a



Desert Spadefoot Toad (*Notaden nichollsi*) and a Wolf Spider (Family: Lycosidae) collected from the pitfall trap (one from morning 1 and two from morning 2). Frogs were only handled for the removal from the trap to prevent damage to their skin from oils (it's best to use gloves for handling where possible).

native Cockroach (Order: Blattodea). The camera traps, funnel traps, Elliott traps, and insect pan traps came up empty, as did the three feral Cat traps. There were, however, plenty of tracks in the sand to observe on both nights. Before setting the traps, some Hopping Mice (*Notomys sp.*) and feral Cat tracks were seen crossing the area. During the trapping, we observed small reptile tracks, frog tracks (easily identified as they led straight to a pitfall that housed the individual) and legless lizard tracks. A set of tracks belonging to a young Australian Bustard was also seen along the road near the survey site, showing that they are in the region.

We also helped to show the Women Rangers how to choose a survey site, as well as show them a range of ways to assess biodiversity from a desktop with a host of excellent online resources (<u>Atlas of Living Australia</u>, <u>NR Maps</u>, <u>NRM InfoNet</u>). We had a great time trapping and look forward to assisting the Tjuwanpa team again in the near future.

We thank the Tjuwanpa Women Rangers and traditional custodians of the land for allowing the Land for Wildlife team to visit and conduct a biodiversity survey. This biodiversity survey was conducted with Animal Ethics approval (Charles Darwin University Animal Ethics 12006 Landscape, fauna and flora survey and impact assessment in relation to mineral and petroleum exploration, infrastructure development and conservation initiatives throughout the Northern Territory), a Parks and Wildlife Commission NT permit (60855 Permit to Interfere with Protected Wildlife) and a Department of Primary Industry and Resources permit (026 Licence to Use Premises for Teaching or Research Involving Animals).

Top: A Pie-dish Beetle (Helea sp.) collected from the pitfall trap on the second morning of trapping and a small reptile seen making its way along the pitfall fence (but diverted away before reaching the bucket). Bottom: Tracks from a small reptile crossing the Elliott/Funnel trap line, tracks from a frog hopping towards a pitfall, a Legless Lizard track near the trapping site, and an Australian Bustard (Ardeotis australis) track on the roadside.





Water is the basis of all life on Earth. Central Australia supports a restricted but significant range of freshwater systems that are used by native plants, animals and humans.

The water found in these freshwater systems varies in age, depending on its origin. Ground water, stored beneath the Earth's surface, accumulates over a long periods of time and may only slowly recharge from rainwater filtering downwards. By contrast, surface water collects from runoff directly following rainfall events.

Understanding the ages of water from individual sources will enable researchers to determine how these valuable natural assets can be most sustainably managed across the landscape. It can also assist with prioritising their conservation to benefit both plants and animals, and the people and enterprises that depend upon them.

### ABOUT THE PROJECT

The Outback Water Project is a collaboration between Territory Natural Resource Management, Charles Darwin University, Parks and Wildlife Commission of the NT and Tourism Central Australia, together with Citizen Scientists.

The project requires people to collect and provide water samples from various waterbodies around the MacDonnell Ranges in Central Australia. These samples will then be analysed by researchers in the environmental lab at Charles Darwin University.

Over time, this information will enable The Outback Water Project to create a map that will indicate the age of the water found in various waterbodies, and provide information on local evaporation rates, humidity, size of rainfall events and the mixing of groundwater and surface water sources.

# **MORE INFORMATION**

For more information about the project, or how to get involved, please see the dedicated Outback Water Project website at: www.outbackwaterproject.com

For updates about the project, and to see some of the sample locations people have visited, see our Facebook page.

Feel free to share your photos or experience collecting a water sample to our Facebook page!



@outbackwaterproject #outbackwaterproject

## **CONTACT US**

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E: jon.hodgetts@territorynrm.org.au













This project is supported through funding from the Australian Government's National Landcare Programme.

## HOW YOU CAN HELP

Citizen Scientists can collect a sample kit from our project hub at the Visitor Information Centre on Alice Springs Todd Mall". These kits will contain rials which can be used to collect water samples from as many locations in the MacDonnell Ranges region as possible.

People are encouraged to also take a photograph of the waterhole that the sample was collected from, to provide visual recordings of the different types of waterholes across the region.

# WHERE YOU CAN HELP

We are seeking water samples from a variety of locations around the MacDonnell Ranges and surrounding areas in Central Australia.

Water samples can be collected from any freshwater body such as water holes, salt takes, soaks, wetlands, clay pans, temporary takes, rock holes, small permanent spring-fed streams and acuiters.

Many of the above waterbodies can be found on publicly accessible areas and are suited to water sample collection.

Please be aware however, that if you intend to take a sample from Aboriginal lands or a private /pastoral property to please obtain permission from the appropriate authority / landholder prior to taking a sample.

#### How to collect and return your sample kit

You can collect and return your sample kit from the following location:

Alice Springs Visitor Information Centre Todd Mall Alice Springs NT

If you are out and about and run into an NT Park Ranger in one of the NT Parks, you can also return your water sample to them.

If you have returned home before getting a chance to drop off your water sample, contact Jon-Hodgetts at jon-hodgetts@territorynrm.org.au, who can advise you on how to submit your water sample.

#### How to collect a water sample

To collect water from your chosen location, go to the water's edge;

- Unscrew lid and fill vial with water from just under the surface
- Try to take as clean a sample as possible i.e. free from significant plant or animal material
- 3. Replace lid
- Fill out label with date of collection, a rough location if known, and GPS co-ordinates if possible (many smart phones have the capability to record GPS when taking a photo. Check the 'details' option usually seen when viewing the photo to see if a GPS location has been recorded.)
- Affix the label to the vial
- Please feel free to take a photo of the site and upload it using the details below

#### How to upload photos of your water collection points

We'd love to see photos of your collection points where you gathered samples. All photos can be uploaded to outbackwaterproject.com.au

Once you reach the website, simply click the 'Submit a photo of your sample site' button in the top right hand comer, then complete and submit the 'Photo Submission Form'.

IMPORTANT: Please make sure you have permission from the landholder BEFORE taking a water sample.

NT Parks are publicly accessible and approved their locations for this project, however samples from Aboriginal lands and private / pastoral properties need express permissions from the relevant authority / landholder



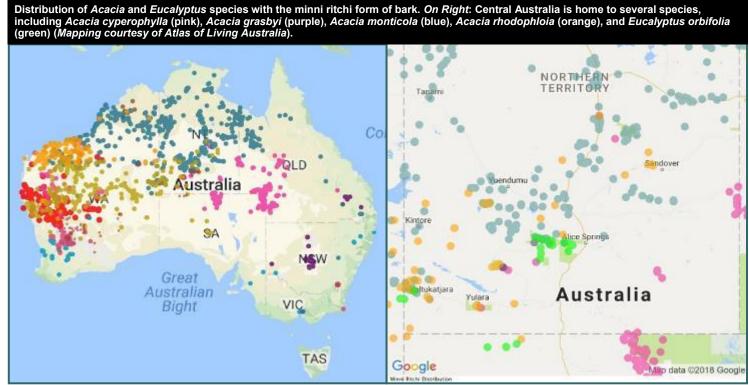
### **Bark Curiosities**

Bark is a non-technical term for the layers of tissue outside the vascular cambium of a tree, woody vine or shrub. The purpose of bark is to protect the tree against sun damage, fire, invertebrates, bacteria and fungi. Bark is made up of two components – the inner bark (living tissue composed of the innermost periderm) and the outer bark (dead tissue on the surface and the outer side of the periderm), also known as rhytidome. The rhytidome is what most people associate with bark, as it is the outer layer that covers the trees and can peel away easily. This outermost layer dies each year and will shed off the trunk in some species, but stay behind for others. This all depends on how the bark of each species responds to the pressure created from the wood on the inside growing and expanding against the outer layers.

There are several types of bark that can be recognised in *Eucalyptus* and closely related groups, which are loosely categorised into smooth bark and rough bark.

#### Smooth Bark

Species that lose their bark annually are the smooth barks, as the rough bark sheds it leaves behind a new layer of smooth living bark. The bark that is shed comes off in slabs, ribbons, or small flakes.





Bark can vary and is shown here in various forms: deeply furrowed cork of the Fork-leaved Corkwood (*Hakea divaricata*), compacted bark on the White Cypress Pine (*Callitris glaucophylla*), both Desert Bloodwood (*Corymbia opaca*) and Ghost Gum (*Corymbia aparrerinja*) have tessellated bark despite the smooth appearance of the latter, an unidentified Western Australian *Eucalyptus sp.* with loose basal slabs, a River Red Gum (*Eucalyptus camaldulensis var. obtusa*) with smooth bark and peeling strips and an Inland Tea-tree (*Melaleuca glomerata*) with paperbark.

(Continued from page 6)

A curious smooth bark is the **minni ritchi**, which is the reason I began looking into bark types! Minni ritchi is a bark that is reddish brown and peels in small curly flakes or fine strips, reminiscent of curly hair. The smooth outer bark splits longitudinally and horizontally, which allows the edges to curl outwards to expose new layers of bark underneath. The name likely derives from an Australian Indigenous language (possibly *minariji*) of unknown origin (*Garuwali*, on the north-east NT border, has been suggested). A number of species of *Acacia* and *Eucalyptus* have minni ritchi bark, of which five have been recorded in central Australia. These include *Acacia cyperophylla*, *Acacia grasbyi*, *Acacia monticola*, *Acacia rhodophloia*, and *Eucalyptus orbifolia*. The species that is well known around Alice Springs is the Red Mulga (*Acacia cyperophylla*), used in many town gardens for its fast-growing nature and ability to survive in many soil types.

#### Rough Bark

Of the rough bark types, there are a few that are classified based on their presence largely in Gums; however the types can extend to other species as well.

**Ironbark** is hard, and becomes compacted and furrowed with age, producing longitudinally furrowed and kino (plant gum) impregnated rough bark that exists over the whole trunk. Colour varies from grey to black and red-black. Local examples include Silver-leaved Ironbark (*Eucalyptus melanophloia*) slightly north-west and Wandi Ironbark (*Eucalyptus jensenii*) in the top end, however the majority of ironbark species are found on the eastern coast.

**Tessellated bark** types are those where the bark splits into flakes or tesserae, which are rough and somewhat corky. In some species, the upper trunk is often smooth, with tessellations becoming more apparent at the basal end of the tree trunk. The weathered tessellations of the Desert Bloodwood (*Corymbia opaca*) are often grey to brown but newly shed flakes leave a yellow to red under layer. While looking somewhat different, the Ghost Gums (*Corymbia aparrerinja*) have tessellated bark that isn't rough in appearance but rather smooth.

**Stringy (or fibrous) bark** encompasses a large group of species that have longitudinally furrowed and fibrous bark that can be pulled off in strings. Some species may appear smooth along the stems and the stringy component found towards the

(Continued from page 7)

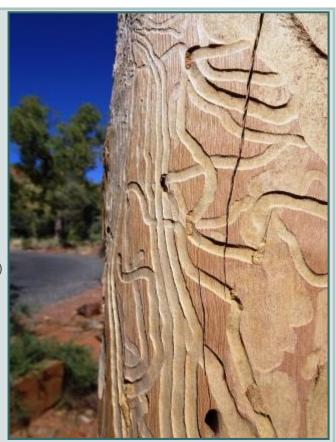
base. While only *Eucalyptus tetrodonta* can be found in the top end, several other species can be found on the eastern coast.

**Box bark** has grey, white or brown short-fibered and tessellated or flaky sections. An example is one of the Coolabah species (*Eucalyptus microtheca*), which can be found locally.

Imperfectly shed ribbons, strips or curls can be found on trees where the bark sheds in either long but coarse ribbons, strips or irregular flakes to expose a smooth surface underneath. This is often confused with loose basal slabs, which is another group altogether (see below).

Loose basal slabs have rough bark, largely on the basal section of the trunk, which accumulates as the bark dies and is held in place Beneath the Bark Lies a Dark Secret

The bark of a tree is designed to help protect the tree from damage (sun, fire, fungus, bacteria, and invertebrates), however some species are adapted to overcome the challenges that bark brings. In central Australia, woodborer beetles are capable of finding their way through tough tree trunks by eating the wood (they are hence termed xylophagous) and include Longhorn Beetles, Bark Beetles, Weevils, and Metallic Flat-headed Borers. They most often attack dying or dead trees and a few species may also become house pests. Here, a Gum at Ormiston Gorge has been attacked by a woodborer.



for a period of time before shedding. This is an ambiguous bark type, as smooth bark on the upper most section of the tree and rough bark on the lower section causes confusion.

**Compacted bark** is similar to ironbark and varies from brown to black, but generally covers the basal region of the trunk, while branches remain smooth. They may often be known as blackbutts.

### More Bark Beyond the Gums

Bark will differ in structure depending on the species, with many developing longitudinal cracks, others plates, flakes and a few will develop horizontal lenticels (which allow for gas exchange). Colouration of the bark will allow for varying degrees of light reflectance, where dark barks absorb light and creamy white bark will reflect it to protect it from sun damage. Some species will allow thick layers of cork to build-up in order to protect the tree from sun damage and fire, whereas thin bark that peels off regularly has the advantage of preventing the accumulation of lichen and other surface damage in moist environments. Bark can also contain chemicals or exude sap to protect the tree from invertebrates, bacteria and fungi. The differences in bark structure are one of the reasons I find them so amazing and wonderful to photograph!

There is little in the way of a standard identification key to bark types outside of the *Eucalyptus* genus, which can lead to confusion in describing a species, though the bark of many species can often be used as a visual cue to identification for those with a little experience or good observation skills. Here are a couple of additional identifiable bark types, but you may find more types when you are out exploring. Feel free to share them with us!

**Corkwood** has thick and heavily furrowed longitudinal cracks. The bark itself is quite light and corky and helps to protect the tree from fire damage. A local example is the Fork-leaved Corkwood (*Hakea divaricata*).

**Paperbark** is a flaky bark that can be pulled off in thin papery strips or flakes. A local example is the Inland Tea-tree (*Melaleuca glomerata*) and others in the *Melaleuca* genus.

#### References:

Brooker, M.I.H. & Kleinig, D.A. (1990). Field Guide to Eucalypts, Volume 2: South-western and Southern Australia. Inkata Press, Melbourne

Australian National Botanic Garden (2018). Euclid. https://www.anbg.gov.au/cpbr/cd-keys/Euclid

American Forests (2013). The Language of Bark. <a href="http://www.americanforests.org/magazine/article/the-language-of-bark/">http://www.americanforests.org/magazine/article/the-language-of-bark/</a>



# Flupropanate Workshop

### By Chris Brown

There is an informal information session on the use of the residual herbicide Flupropanate for the control of Buffel Grass (*Cenchrus ciliaris*) and Couch Grass on May 4th at the Desert Knowledge Precinct from 10:00 AM to 12:00 PM.

Flupropanate is a residual herbicide used for the control of a variety of perennial grasses including Serrated Tussock, Buffel Grass and Couch Grass. The advantages of Flupropanate is that it can be applied to Buffel Grass when it is not actively growing, greatly expanding the control window, it also provides control of residual seedbanks as well as parent plants, and provides long lasting control up to 2 years in some locations such as SA (unlike glyphosate which can only be applied when plants are actively growing with no lasting control of soil borne seedbanks).

Flupropanate has been used extensively in South Australia (where Buffel Grass is a declared weed) with great success (see SA PIRSA herbicide trial report). Trials are currently underway in Central Australia at several locations including the Desert Knowledge Precinct which will form part of the information session. Bill Dobbie who is the man behind the development of Taskforce, the registered trade name for Flupropanate in Australia, is willing to deliver an information session on how to get the most benefit out of using Flupropanate for the control of Buffel Grass and Couch Grass. As well as having a scientific background, Bill is also a landholder with firsthand experience gained over several decades.

Can everyone please let me [Chris] know ASAP of their interest / availability in attending the information session by emailing chris.brown@nt.gov.au or calling (Ph 08 8951 9210, Mob 0458 515 243).

#### ~ Chris Brown

(Regional Weed Officer, Weed Management Branch, Department of Environment and Natural Resources, Northern Territory Government)



L to R: A Waddy Wood (Acacia peuce) being presented to Mayor Smith by Mr John Blakeman in 1981 (Image: The Star); the tree in 1989 as a small sapling (Image: Land for Wildlife); and in 1999 (Image: Land for Wildlife). This tree is now listed in the Alice Springs region of the NT Register of Significant Trees #24.

# A Snippet From Significant Trees

### » Alice Springs Town Council's Waddy Wood

The Waddy Wood (*Acacia peuce*) found on the Alice Springs Town Council lawns is one of the rarest (vulnerable EPBC listing and found in only three locations) and most striking trees of the Australian arid zone. This particular individual was planted on World Environment Day in 1981 after it was presented to Mayor George Smith by John Blakeman MBE. It was presented on behalf of the Society for Growing Australian Plants Alice Springs, who had grown it from seed collected on Andado Station. A mix-up occurred behind the scenes which led to some red faces and some stealthy activity in the night, but to find out more you will have to attend a walking tour by the Land for Wildlife team! It has been listed in the NT Register of Significant Trees since 1996 for cultural reasons and its rare nature.

View the <u>NT Register of Significant Trees</u> page to learn more about the register. *The NT Register of Significant Trees was established by the National Trust NT and Greening Australia and is managed by Land for Wildlife Central Australia.* 



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# Significant Trees Walking Tour

Candice Appleby from Land for Wildlife ran a walking tour of some of the CBD's most significant trees this month as part of the National Trust Heritage Festival. The tour went on a moderately paced 1 Km walk from the Hartley Street School to Wills Terrace via the ASTC lawns and Todd Mall.

The walking tour was attended by twelve enthusiastic history buffs, who were eager to learn about the stories behind some of the listings on the NT Register of Significant Trees. We were excited to have Alex Nelson and Stuart Traynor come along to add some extra pieces of the puzzle—resulting in a lively and organic mix of information. Thanks to all who attended and contributed to the discussion!



# Soil Crusting and Cryptograms

The ecological significance of biological soil crusting and the presence of cryptograms has been outlined in a brand new fact sheet. You may remember the November 2007 Newsletter that addressed this—download the updated fact sheet to read about why soil crusting is so important on your property.

### **APS Plant Sale**

The Australian Plants Society Alice Springs Inc had their biannual plant sale recently and Land for Wildlife was there to promote the program and provide assistance with vegetation mapping. We had some newly interested visitors looking to sign up and a few existing members stopped by to say hi. The sale itself was a massive success, with queues making their way through the Olive Pink Botanic carpark. Well done, guys!

Are you interested in becoming an APS Member? You can find their details below. Get in touch to sign up, head along to one of their meetings and find out about other local groups of interest on the Land for Wildlife <u>Networks</u> webpage.

Australian Plants Society Alice Springs Inc: An enthusiastic group of with a primary interest in learning about the local native plants. Meetings take place at Olive Pink Botanic Garden, Tuncks Road once a month. They are also involved in other activities including a display at the annual Alice Springs Show, native plant sales, monitoring populations of rare plants and field trips to observe and learn about unique central Australian flora.

- w. Australian Plants Society Alice Springs Inc.
- e. apsalicesprings@yahoo.com.au
- p. 08 8952 2154



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A Garden for Wildlife member, Rosalie, has sent us in this photo of a Bearded Dragon (*Pogona vitticeps*) found in her garden at East Side. What a great find for an urban garden! Well done, Rosalie! (*Image R. Schultz*)



MEET ON THE LAST SUNDAY OF EACH MONTH FROM 7:30AM TO 12PM TO HELP OUR BEAUTIFUL ALICE SPRINGS' BIODIVERSITY\*

SUNDAY APR 29

SPENCER VALLEY BUFFEL CONTROL

CALL SUE 0409412045 SUNDAY MAY 27

ILPARPA CLAYPANS BUFFEL CONTROL

> CALL MARIE 0467879254

SUNDAY Jun 24

KURRAJONG DRIVE BUFFEL CONTROL

> CALL VALMAI 0488033574

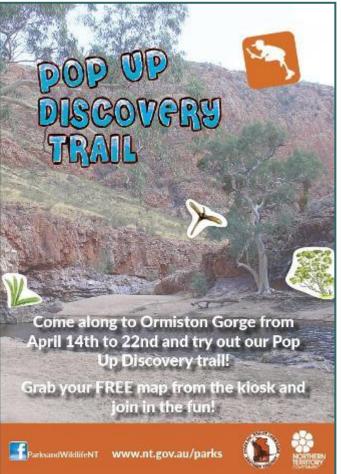
SUNDAY JUL 29

TODD RIVER Buffel Control

> CALL KEN 0413596012

\*PLEASE WEAR COVERED FOOTWEAR AND BYO HAT, WATER, GLOVES AND A MATTOCK IF YOU HAVE ONE!

INFO@ALICESPRINGSLANDCARE.COM | FB.COM/ASLANCARE | WWW.ALICEPSINRGSLANDCARE.COM





# AGM Notice and Call for nominations

May 16<sup>th</sup> 2018

Birdlife Central Australia will hold its Annual General Meeting on Wednesday May 18<sup>th</sup>, 2018 at the Alice Springs Desert Park Meeting Room on Larapinta Dr, Alice Springs commencing at 7pm.

#### Agenda:

- 1. Apologies
- 2. Annual reports from office bearers
- 3. Election of all committee positions
- 4. Other business

#### **Nominations for Committee Members**

Nominations are called for all positions on the committee for a one year term commencing 17 May 2018. Nominations should be received by the secretary 28 days prior to the AGM.

Birdlife Central Australia members wishing to nominate should complete the attached nomination form and return to Pam Walker, BirdLife Central Australia, 31 Clarke St Araluen NT 0870 by 5:00 pm, 18th April 2018. A scanned copy of a signed nomination form emailed to <a href="mailto:birdlifeca@gmail.com">birdlifeca@gmail.com</a> by 5:00 pm, 18th April 2018 will also be accepted.

Nominations received will be emailed to all members.

If the number of nominations exceeds one for each position, an election will be held at the AGM on 17 May 2018. If an insufficient number of nominations are received to form a complete committee, nominations will be called for from the floor at the AGM.



Further Reading
Click the link symbol
to be redirected to the article



Article • Maligned dingoes emerge as unlikely heroes in study of bushfires and feral pests



Article • Stick-nest rats are masters of construction



Article • Scientists crack the code of shell strength, paving the way for safer eggs



Article • This fat little circle builds a nest to die for



Article • Birds can see Earth's magnetic fields, and we finally know how that's possible

#### Cheers,

# Caragh, Candice and Bill

Do you have any stories or images to share? Get in touch! We are always looking for members to share their experiences via our social media and newsletter. Email us with your suggestions of articles or topics that you wish to hear more about.

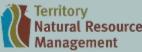
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Northern Territory
Government





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## **Contact Us**

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